AMENDMENTS TO THE CLAIMS

1-8. (canceled)

- 9. (currently amended) A method for real-time detecting and quantifying a first nucleic acid template and a second nucleic acid template in a PCR mixture comprising the steps of
- thermally cycling a PCR mixture wherein the PCR mixture comprises a thermostable polymerase, a double stranded DNA dye, the first template and the second template, primers for amplifying a first amplicon from the first template and a second amplicon from the second template, and wherein the first amplicon has a first T_m and the second amplicon has a second T_m and the first T_m is less than the second T_m ;
- b) obtaining cycle by cycle a first emission reading of the double stranded

 DNA dye at a first MT measuring temperature between an annealing/extension

 temperature and the first T_m and a second emission reading of the double stranded

 DNA dye at a second MT measuring temperature between the first T_m and the second T_m;
- c) determining cycle by cycle a first emission amount of the first amplicon which is the difference between the first emission reading and the second emission reading, and a second emission amount of the second amplicon which is the second emission reading.

10. (canceled)

- 11. (original) The method of claim 9 wherein the double stranded DNA dye is a double stranded DNA intercalating dye.
- 12. (original) The method of claim 11 wherein the double stranded DNA intercalating dye is selected from the group consisting of ethidium bromide, YO-PRO-1, Hoechst 33258, SYBR Gold, and SYBR Green I.
- 13. (original) The method of claim 9 wherein the double stranded DNA dye is a primer-based double stranded DNA dye.
- 14. (original) The method of claims 13 wherein the primer-based double stranded DNA dye is selected from the group consisting of fluorescein, FAM, JOE, HEX, TET, Alexa Fluor 594, ROX, and TAMRA, rhodamine, BODIPY-FI.
- 15. (currently amended) The method of claim 9 wherein the first $\frac{MT}{measuring}$ temperature is 0.25 O C below the first T_m , 0.5 O C below the first T_m , 1.0 O C below the first T_m , 1.5 O C below the first T_m , or 2.0 O C below the first T_m , and wherein the first $\frac{MT}{measuring}$ temperature is higher than the annealing temperature.
- 16. (currently amended) The method of claim 9 wherein the second $\frac{MT}{measuring \ temperature}$ is 0.25 ^{O}C below the second T_m , 0.5 ^{O}C below the second T_m , 1.0 ^{O}C below the second T_m , 1.5 ^{O}C below the second T_m , or 2.0 ^{O}C below the second T_m , and wherein the second $\frac{MT}{measuring \ temperature}$ is higher than the first T_m .
- 17. (currently amended) The method of claim 9 wherein the second $\frac{MT}{measuring temperature}$ is 0.25 ^{O}C above the first T_m , 0.5 ^{O}C above the first T_m , 1.0 ^{O}C above the first T_m , 1.5 ^{O}C above the first T_m , or 2.0 ^{O}C above the first T_m , and wherein the second $\frac{MT}{m}$ measuring temperature is less than the second T_m .

18. (currently amended) The method of claim 9 wherein the second MT measuring temperature is the first $T_m + 0.25^{\circ}C$ < the second MT measuring temperature < the second T_m -0.25°C, the first $T_m + 0.5^{\circ}C$ < the second MT measuring temperature < the second T_m -0.5°C, the first $T_m + 1.0^{\circ}C$ < the second MT measuring temperature < the second T_m -1.0°C, the first $T_m + 1.5^{\circ}C$ < the second MT measuring temperature < the second T_m -1.5°C, or the first $T_m + 2.0^{\circ}C$ < the second MT measuring temperature < the second T_m -2.0°C.

19. (canceled)

- 20. (currently amended) The method of claim 9 wherein the <u>first</u> emission amount of the first amplicon is obtained through a computer program performing a calculation of subtracting the first emission <u>reading</u> from the second emission <u>reading</u> or subtracting the second emission <u>reading</u> from the first emission <u>reading</u>.
- 21. (currently amended) A method for real-time detecting and quantifying a first nucleic acid template and a second nucleic acid template in a PCR mixture comprising the steps of:
 - a) thermally cycling a PCR mixture wherein the PCR mixture comprises a thermostable polymerase, a double stranded DNA dye, the first template and the second template, primers for amplifying a first amplicon from the first template and a second amplicon from the second template, and wherein the first amplicon has a first T_m and the second amplicon has a second T_m and the first T_m is less than the second T_m;
 - b) obtaining cycle by cycle a first pre-T_m emission reading of the double stranded DNA dye at a MT measuring temperature below the first T_m and a first post-T_m emission reading of the double stranded DNA dye at the a MT measuring temperature above the first T_m and a second pre-T_m emission reading of the double stranded DNA dye at a MT

- <u>measuring temperature</u> below the second T_m and a second post- T_m emission <u>reading of the double stranded DNA dye</u> at the a MT measuring temperature above the second T_m ;
- c) determining cycle by cycle a first emission amount of the first amplicon which is the difference between the first pre-T_m emission <u>reading</u> and the first post-T_m emission <u>reading</u>; and a second emission amount of the second amplicon which is the difference between the second pre-T_m emission <u>reading</u> and the second post-T_m emission <u>reading</u>.
- 22. (original) The method of claim 21 wherein the double stranded DNA dye is a double stranded DNA intercalating dye
- 23. (original) The method of claim 22 wherein the double stranded DNA intercalating dye is selected from the group consisting of ethidium bromide, YO-PRO-1, Hoechst 33258, SYBR Gold, and SYBR Green I.
- 24. (original) The method of claim 21 wherein the double stranded DNA dye is a primer-based double stranded DNA dye.
- 25. (original) The method of claims 24 wherein the primer-based double stranded DNA dye is selected from the group consisting of fluorescein, FAM, JOE, HEX, TET, Alexa Fluor 594, ROX, and TAMRA, rhodamine, BODIPY-FI.
- 26. (currently amended) The method of claim 21 wherein the $\frac{MT}{measuring}$ temperature below the first T_m and/or the second T_m are 0.25 ^{O}C below, 0.5 ^{O}C below, 1.0 ^{O}C below, 1.5 ^{O}C below, or 2.0 ^{O}C below.
- 27. (currently amended) The method of claim 21 wherein the $\frac{MT}{m}$ measuring temperature above the first T_m and/or the second T_m are 0.25 O C above, 0.5 O C above, 1.0 O C above, 1.5 O C above, or 2.0 O C above.

28. (currently amended) The method of claim 21 wherein the <u>first_emission</u> amount of the <u>first_amplicons</u> is obtained through a computer program performing the calculation of subtracting the <u>first_pre-T_m</u> emission <u>reading</u> from the <u>first_post-T_m</u> emission <u>reading</u> from the <u>first_pre-T_m</u> emission <u>reading</u>, and the second emission amount of the second amplicon is obtained through the computer program performing the calculation of subtracting the second <u>pre-T_m_emission_reading_or_emission_re</u>

29-84. (canceled)

- 85. (new) A method for real-time detecting and quantifying a first nucleic acid template and a second nucleic acid template in a PCR mixture comprising the steps of
- a) thermally cycling a PCR mixture wherein the PCR mixture comprises a thermostable polymerase, a double stranded DNA dye, the first template and the second template, primers for amplifying a first amplicon from the first template and a second amplicon from the second template, and wherein the first amplicon has a first T_m and the second amplicon has a second T_m and the first T_m is less than the second T_m;
- b) obtaining cycle by cycle a first emission reading of the double stranded DNA dye at a first measuring temperature between an annealing/extension temperature and the first T_m , a second emission reading of the double stranded DNA dye at a second measuring temperature between the first T_m and the second T_m ; and a third emission reading of the double stranded DNA dye at a third

measuring temperature between the second T_m and a total denaturing temperature; and.

- c) determining cycle by cycle a first emission amount of the first amplicon which is the difference between the first emission reading and the second emission reading, and a second emission amount of the second amplicon which the difference between the second emission reading and the third emission reading.
- 86. (new) The method of claim 85 wherein the double stranded DNA dye is a double stranded DNA intercalating dye.
- 87. (new) The method of claim 86 wherein the double stranded DNA intercalating dye is selected from the group consisting of ethidium bromide, YO-PRO-1, Hoechst 33258, SYBR Gold, and SYBR Green I.
- 88. (new) The method of claim 85 wherein the double stranded DNA dye is a primer-based double stranded DNA dye.
- 89. (new) The method of claims 88 wherein the primer-based double stranded DNA dye is selected from the group consisting of fluorescein, FAM, JOE, HEX, TET, Alexa Fluor 594, ROX, and TAMRA, rhodamine, BODIPY-FI.
- 90. (new) The method of claim 85 wherein the first measuring temperature is 0.25 O C below the first T_m , 0.5 O C below the first T_m , 1.0 O C below the first T_m , or 2.0 O C below the first T_m , and wherein the first measuring temperature is higher than the annealing temperature.

- 91. (new) The method of claim 85 wherein the second measuring temperature is $0.25\,^{\circ}\text{C}$ below the second T_m , $0.5\,^{\circ}\text{C}$ below the second T_m , $1.0\,^{\circ}\text{C}$ below the second T_m , $1.5\,^{\circ}\text{C}$ below the second T_m , or $2.0\,^{\circ}\text{C}$ below the second T_m , and wherein the second measuring temperature is higher than the first T_m .
- 92. (new) The method of claim 85 wherein the second measuring temperature is 0.25 $^{\circ}$ C above the first T_m , 0.5 $^{\circ}$ C above the first T_m , 1.0 $^{\circ}$ C above the first T_m , or 2.0 $^{\circ}$ C above the first T_m , and wherein the second measuring temperature is less than the second T_m .
- 93. (new) The method of claim 85 wherein the second measuring temperature is the first $T_m + 0.25^{\circ}C$ < the second measuring temperature < the second T_m -0.25°C, the first $T_m + 0.5^{\circ}C$ < the second measuring temperature < the second T_m -0.5°C, the first $T_m + 1.0^{\circ}C$ < the second measuring temperature < the second T_m -1.0°C, the first $T_m + 1.5^{\circ}C$ < the second measuring temperature < the second T_m -1.5°C, or the first $T_m + 2.0^{\circ}C$ < the second measuring temperature < the second T_m -2.0°C.
- 94. (new) The method of claim 85 wherein the third measuring temperature is 0.25 O C above the second T_m , 0.5 O C the second T_m , 1.0 O C above the second T_m , and wherein the third measuring temperature is less than the total denaturing temperature.
- 95. (new) The method of claim 85 wherein the first emission amount of the first amplicon is obtained through a computer program performing a calculation of subtracting the first emission reading from the second emission reading or subtracting the second emission reading from the first emission reading, and the second emission amount of the second amplicon is obtained through a computer program performing a calculation of subtracting the second emission reading from the third emission reading or subtracting the third emission reading from the second emission reading.

96. (new) The method of claim 21 wherein the measuring temperature above the first T_m and the measuring temperature below the second T_m is the same.